



MARINE CARBON SENSING WORKSHOP:

Transformative full-ocean
depth sensor platforms

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Current in situ sensor platforms

DRIFTER

Cheap and easily deployed, power in numbers

Cost: \$500-6,000
Depth: 0-100m
Motion: Lagrangian drift

Ex: Sonobuoys, Sofar Spotter

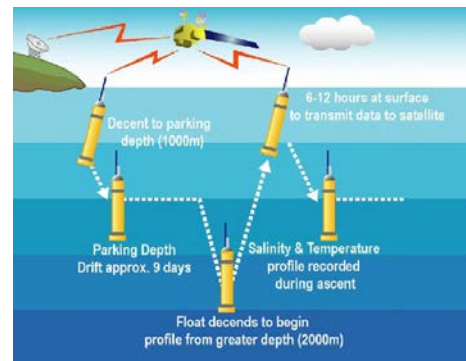


FLOAT (profiler)

Ability to change depth by adjusting buoyancy or crawling a mooring line

Cost: \$20,000-100,000
Depth: 100-6000m
Motion: Moored or drifting

Ex: Argo program

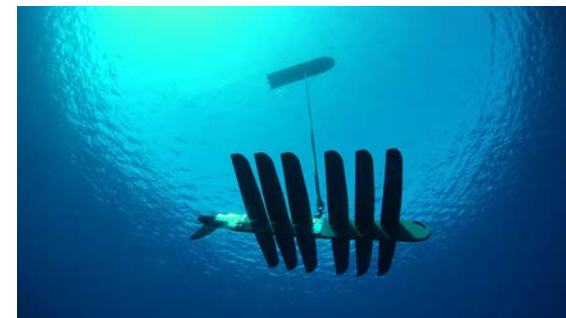


AUTONOMOUS

Self-powered, steerable, long activity cycle

Cost: \$200,000-10,000,000
Depth: 200-1,000m though some have reached 6000m
Motion: Pseudo-active

Ex: Wave Glider



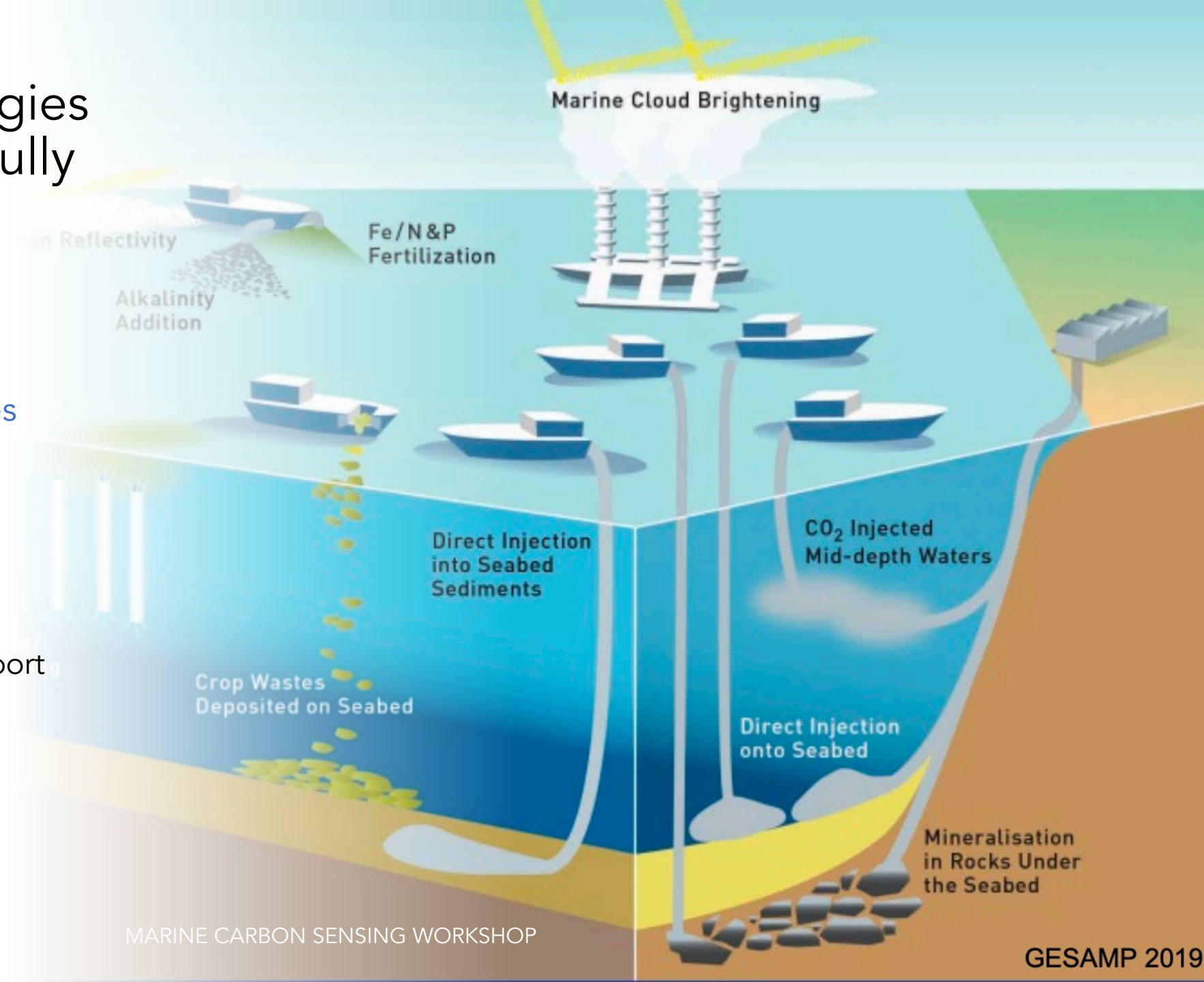
Platform technologies critical to successfully measuring and reporting of CO₂

- Measurement Challenges

- Spatial
 - $10^{-2} \rightarrow 10^2$ km
- Temporal
 - $10^{-3} \rightarrow 10^2$ years

- Platform Challenges

- Environment
- Efficiency/ Cost of Transport
 - Range
- Energy generation
 - Duration
- Robustness
- Economics

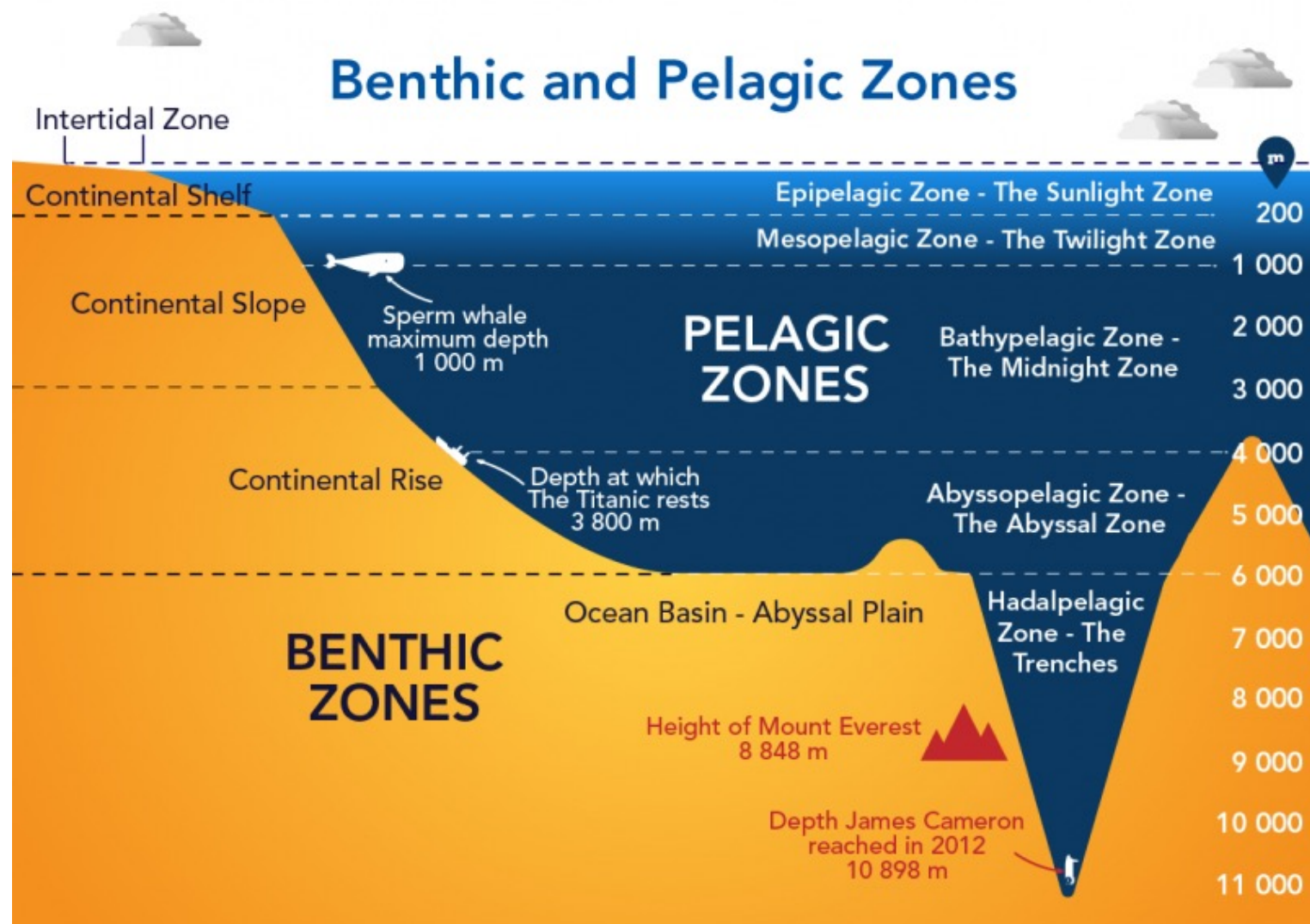


Ability to actively/passively monitor carbon levels in all ocean zones will require new solutions in the underwater platform design space

Opportunity to go beyond traditional platform design and take inspiration from biology: manmade underwater vehicles have had ~250 years to evolve, biology... millions of years

DIVERSITY OF DESIGN:
35,000+ species of known
fishes occupying all
oceanic zones presents
incredible morphological
diversity

Mother
nature
offers
inspiration



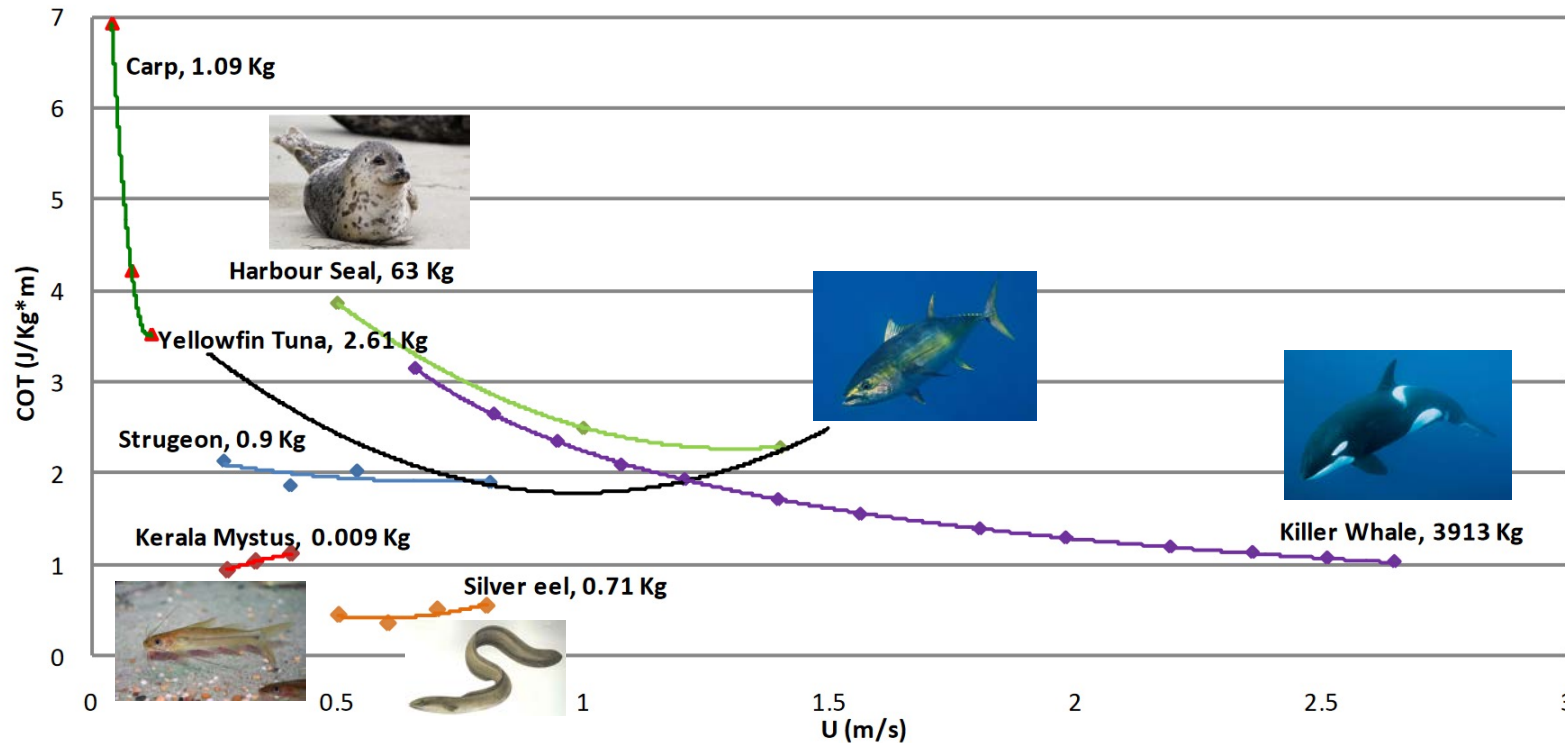
Biological Propulsion: Diversity of Design & Inspiration

- **Biology has evolved solutions to inhabit all oceanic zones**
 - Offers the potential for game changing solutions for autonomous platform designs that form the framework to quantifying carbon in the ocean
 - Must be clear on what aspects of their solution are critical to success
 - SPATIAL/TEMPORAL//ENERGETICS/COST/ETC CONSTRAINTS
 - Idea is not to mimic biology but be inspired
- **Fishes are not pressure vessels**
 - IDEA OF DEPTH AGNOSTIC SYSTEMS
 - Removing pressure vessel design through soft, solid-state components opens up deep ocean zones for MRV
 - ECONOMICS OF SOFT SYSTEMS
 - Cost less than traditional *metal-foam* UUV designs

Cost of Transport

Quantifying Energy Efficiency

Cost of Transport (i.e., energy efficiency): varies with speed and species



- Active metabolic rate of fish is analogous to electrical power consumption of platform ==> cost of transport scaled by body mass (J/kg/m) **allows biology-robot comparison directly**
- CoT is a key metric to quantify
- Understand recharging requirements for long duration sensing runs
 - *Charging stations (analogous to cleaning stations of Manta Rays)*

(Phillips et al., 2012, Further Advances in Unmanned Marine Vehicles, edited by G.N. Roberts and R. Sutton ISBN: 978-1-84919-479-2)

Pressure drag



Lift-based propulsion



Acceleration reaction

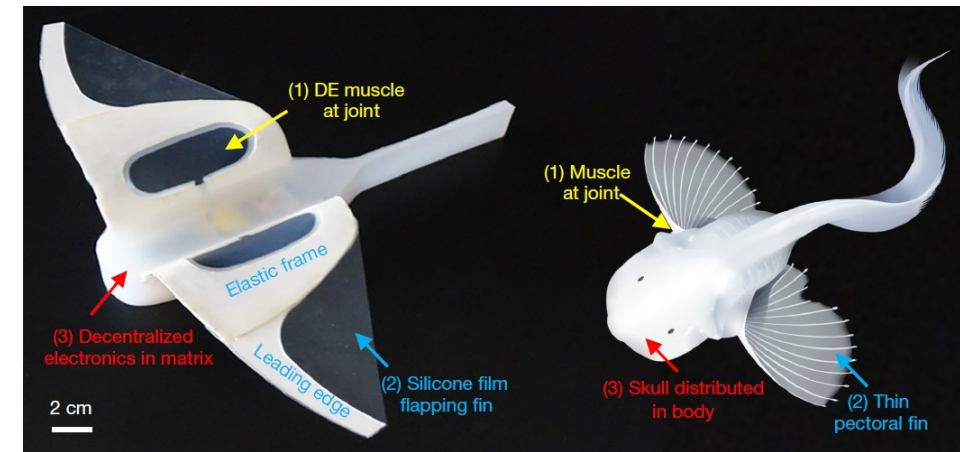


Biological Propulsion:
Diversity of Design & Inspiration

Lift-based propulsion

GhostSwimmer Autonomous Undersea Vehicle (AUV) by Boston Engineering

- Tuna-inspired
- Maximum depth: 100 m
- Endurance: 14 hrs



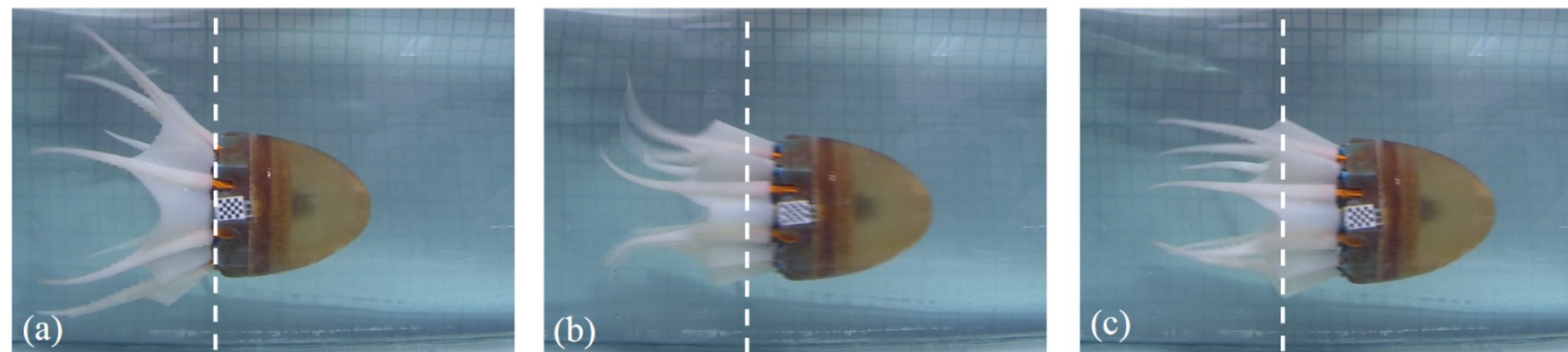
Soft robot (left) inspired by snailfish (right)

- Swam at depth of 10,900 m in the Mariana Trench
- Actuated by dielectric elastomer (DE) material
- Li et al. Self-powered soft robot in the Mariana Trench, Nature (2021) doi: 10.1038/s41586-020-03153-z

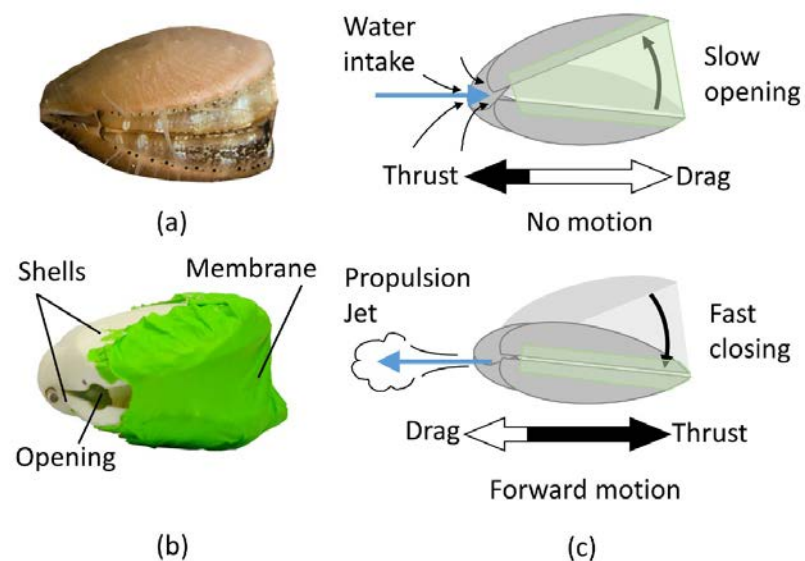


- Cai et al., "Research on Robotic Fish Propelled by Oscillating Pectoral Fins", Robot Fish, 2015 ISBN:978-3-662-46870-8

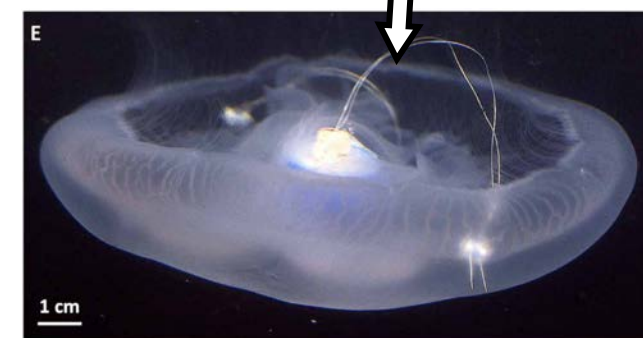
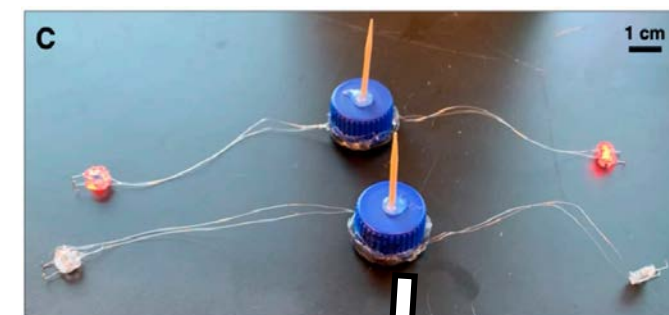
Acceleration reaction



Soft, compliant web made of silicone
Sfakiotakis et al. (2014) doi:10.1109/IROS.2014.6942576



- “RoboScallop”
- Powered by a single DC motor
- Robertson et al. (2019)
doi:10.1109/LRA.2019.2897144



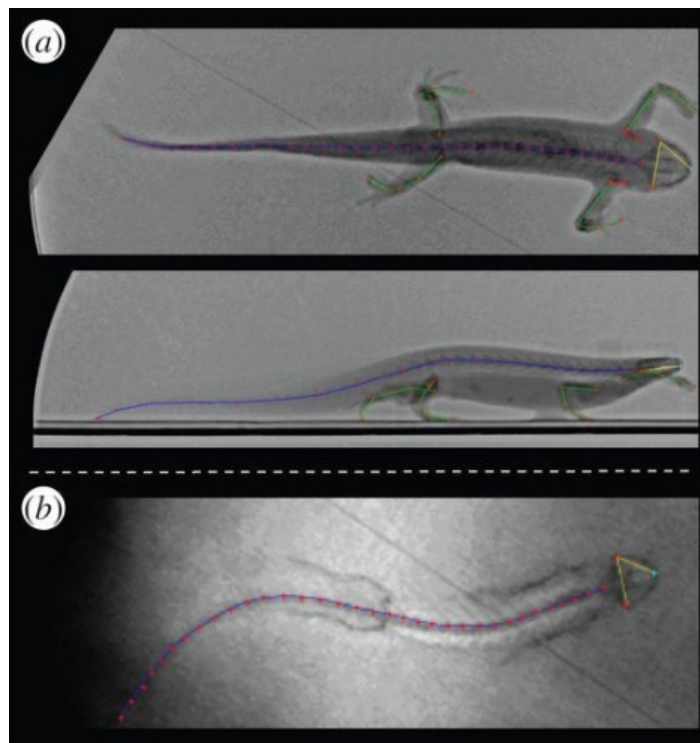
Low-power microelectronics embedded in live jellyfish enhance propulsion
(Xu, ..., **Dabiri** (2020) doi:10.3390/biomimetics5040064)
(Xu & **Dabiri** (2020) doi:10.1126/sciadv.aaz3194)

Pressure drag

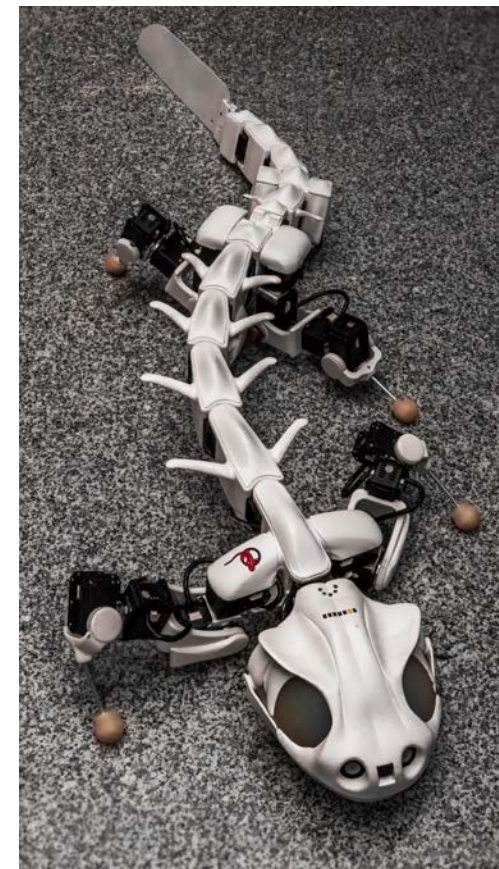


- Turtle-inspired robot using soft material flippers change shape into legs
- Rebecca Kramer-Bottiglio (Yale University), Frank Fish (West Chester University), Simon Freeman (NUWC) (<https://www.youtube.com/watch?v=Q9FyaRtLOys>)

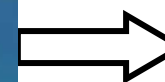
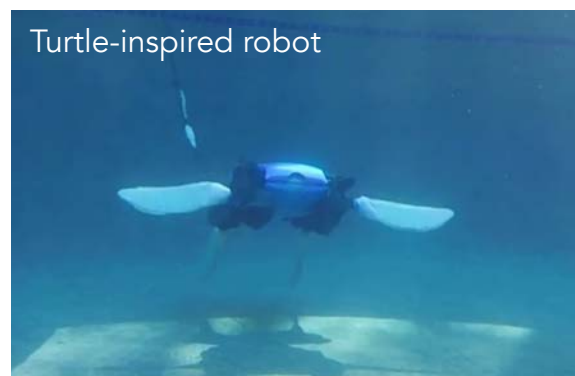
Amphibious Systems: Land & Sea-Based Missions



"Pleurobot": salamander-like robot
Karakasiliotis, Thandiackal, et al. (2016), From cineradiography to biorobots: an approach for designing robots to emulate and study animal locomotion, Royal Society Interface
doi:10.1098/rsif.2015.1089



Turtle-inspired robot



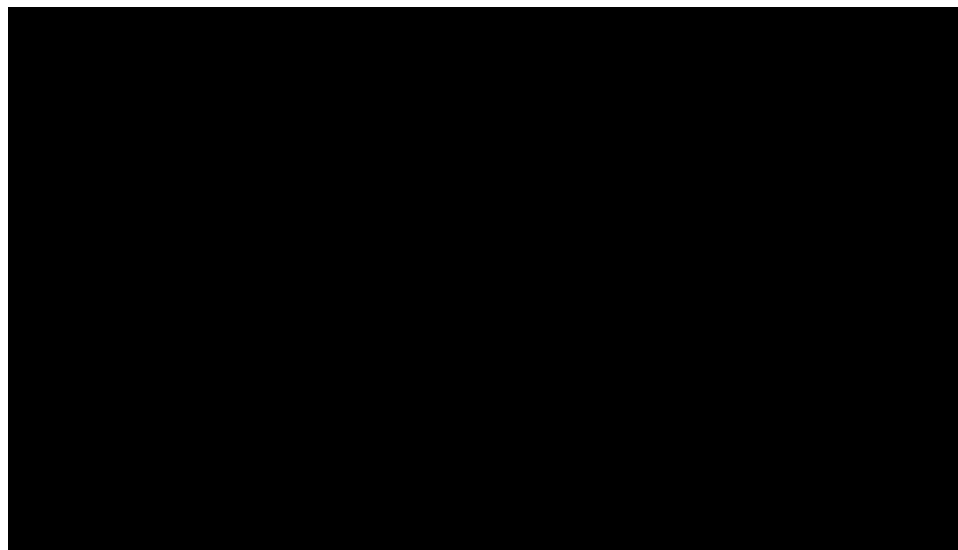
Speed and Cost of Transport: Literature Review

Reference	Description	Speed (m/s)	Body Length BL (m)	Speed (BL/s)	Actuation Frequency (Hz)
Andersen & Chhabra (2002)	VOUTV	1.25	2.4	0.52	1.0
Barrett et al. (1999)	Rabatuna	0.70	1.25	0.56	1.1
Berlinger et al. (2021)	Finbot	0.122	0.160	0.763	4.25
Bujard et al. (2021)	Robotic squid	0.26	0.266	0.98	9.0
Butail et al. (2015)	Robotic zebrafish	0.04	0.154	0.26	3.0
Cai et al. (2010)	Raba-ray II	0.157	0.32	0.49	1.2
Cai et al. (2015)	Robotic caunare ray	0.30	0.46	0.65	0.6
Chen & Jia (2019)	Tensegrity robotic fish	0.30	0.420	0.72	1.72
Chen et al. (2015)	IPMC robotic cownose ray	0.007	0.21	0.034	0.157
Chen et al. (2019)	IPMC robotic fish	0.12	0.27	0.45	1.0
Chen et al. (2021)	Leaping fish bot	1.88	0.264	7.12	15.04
Christianson et al. (2018)	DEA robotic leptocephali	0.0019	0.22	0.0086	0.33
Christianson et al. (2019)	DEA robotic jellyfish	0.0032	0.163	0.020	0.2
Cianchetti et al. (2015)	OCTOPUS	0.05	0.17	0.3	0.75
Curet et al. (2011)	Robotic knife fish	0.30	0.459	0.65	6.0
Du et al. (2019)	Carifuna	0.80	0.52	1.52	5.0
Erturk (2015)	MFC piezoelectric fish	0.075	0.243	0.31	5.0
Faridodin Hosseini et al. (2014)	UC-like I	0.29	0.70	0.41	3.0
Fish et al. (2017)	MantaBot	0.43	0.428	1.00	1.1
Fujiwara & Yamaguchi (2017)	Single-motor-actuated fish	0.58	0.345	1.7	16.0
Gilman et al. (2018)	Flexible robotic fish	0.104	0.17	0.61	1.6
Hirata et al. (2000)	Robotic zebrafish	0.20	0.34	0.59	2.3
Juraffi et al. (2017)	Pneumatic swimmer	0.13	0.17	0.75	0.55
Katzschmann et al. (2018)	Safi	0.235	0.47	0.50	1.4
Kumpf (2000)	Robopike	0.09	0.82	0.1	1.0
Kwak & Bae (2017)	Robotic water beetle	0.117	0.095	1.2	1.3
Loftwich & Smith (2011)	Robotic lamprey	0.115	0.14	0.85	1.0
Loftwich et al. (2012)	Robotic lamprey	0.10	0.90	0.11	0.56
Liu et al. (2013)	Underactuated robotic fish	0.15	0.425	0.35	1.0
Liu et al. (2017)	Electro-ionic robotic fish	0.135	0.093	1.5	5.0
Liu et al. (2019)	Group of robotic fish	0.575	0.45	1.28	1.4
Liu et al. (2021)	Mariana Trench zebrafish	0.0519	0.22	0.24	1.0
Liu et al. (2014)	Wire-driven robotic fish	0.333	0.495	0.67	1.0
Liu & Curet (2018)	KnifeBot	0.325	0.462	0.703	3.0
Luo et al. (2006)	BEAR swimmer	0.049	0.32	0.21	1.7
Luo et al. (2006)	Medicine	0.74	0.75	0.95	6.0
Mazlan (2015)	Robosalmon	0.143	0.90	0.16	1.0
Mazumdar et al. (2009)	Compliant Robotic Tuna (CRT)	0.10	0.27	0.37	2.0
McGovern et al. (2009)	NEMO-propelled fish	0.033	0.125	0.26	0.8
Mahmoudkhaki et al. (2008)	ADCSL robotic fish	0.75	0.6	1.25	4.0
Neely et al. (2016)	Robotic stingray	0.094	0.35	0.27	1.4
Parshat et al. (2017)	Reconfigurable armed robot	0.1	0.6	0.2	2.0
Pham et al. (2019)	Pectoral fin-propelled fish	0.231	0.4	0.58	0.75
Roberson et al. (2019)	RabaScallap	0.16	0.8	0.20	2.56
Sfakiotakis et al. (2015)	Robotic octopus	0.0986	0.38	0.26	0.9
Shintake et al. (2018)	DEA robotic fish	0.0372	0.15	0.25	0.75
Shintake et al. (2020)	Tensegrity trout robot	0.23	0.400	0.58	3.0
Villanueva et al. (2011)	Robojelly	0.0542	0.6	0.1	0.5
Villanueva et al. (2013)	Cyro	0.0847	0.316	0.268	0.12
Wang et al. (2010)	SPC-3 UUV	1.87	1.6	1.2	2.5
Wang et al. (2021)	Robotic larval zebrafish	0.133	0.0043	31	4.8E-05
White et al. (2020)	Tunabot Flex	1.17	0.255	4.60	3.6E-04
White (2022)	Tunabot Prototype	0.64	0.4064	1.6	10.8
Yu et al. (2016a)	Single-motor-actuated fish	1.14	0.37	3.1	16.9
Zhong et al. (2017)	Wire-driven robotic fish	0.67	0.31	2.2	22.5
Zhu et al. (2019)	Tunabot	1.02	0.255	4.00	5.6

Reference	Description	Speed (m/s)	Body Length, BL (m)	Speed (BL/s)	Actuation Frequency (Hz)	Power (W)	Work per Meter (J/m)	Mass (kg)	COT (J/kg/m)
Berlinger et al. (2021)	Finbot	0.122	0.160	0.763	4.25	3.3	27	0.15	180
Bujard et al. (2021)	Robotic squid	0.26	0.266	0.98	9.0	0.09	0.4	0.380	0.93
Chen et al. (2015)	IPMC robotic cownose ray	0.007	0.21	0.034	0.157	2.0	280	0.119	2354
Chen et al. (2021)	Leaping fish bot	1.88	0.264	7.12	15.04	89	47	0.350	135
Christianson et al. (2018)	DEA robotic leptocephali	0.0019	0.22	0.0086	0.33	0.020	10.5	0.0251	419
Christianson et al. (2019)	DEA robotic jellyfish	0.0032	0.163	0.020	0.2	0.25	78	0.23	340
Cianchetti et al. (2015)	OCTOPUS	0.05	0.17	0.3	0.75	2.6	53	3.0	18
Erturk (2015)	MFC piezoelectric fish	0.075	0.243	0.31	5.0	1.4	19	n/a	n/a
Fujiwara & Yamaguchi (2017)	Single-motor-actuated fish	0.58	0.345	1.7	16.0	20.4	35	0.597	59.0
Kumpf (2000)	Robopike	0.09	0.82	0.1	1.0	8.5	94	n/a	n/a
Kwak & Bae (2017)	Robotic water beetle	0.117	0.095	1.2	1.3	0.66	5.7	0.02265	250
Li et al. (2017)	Electro-ionic robotic fish	0.135	0.093	1.5	5.0	0.024	0.18	0.0425	4.18
Liu & Curet (2018)	KnifeBot	0.325	0.462	0.703	3.0	2.55	7.85	n/a	n/a
Long Jr. et al. (2006)	Madeleine	0.74	0.78	0.95	6.0	58.3	79	24.4	3.2
Mazlan (2015)	Robosalmon	0.143	0.90	0.16	1.0	5.4	37.8	4.30	8.8
Paschal et al. (2017)	Reconfigurable armed robot	0.1	0.6	0.2	2.0	4.51	45	2.1	21
Pham et al. (2019)	Pectoral fin-propelled robot	0.231	0.4	0.58	0.75	0.102	0.44	1.059	0.42
Sfakiotakis et al. (2015)	Robotic octopus	0.0986	0.38	0.26	0.9	3.83	38.8	2.68	14.5
Shintake et al. (2018)	DEA robotic fish	0.0372	0.15	0.25	0.75	0.92	25	0.0044	5621
Shintake et al. (2020)	Tensegrity trout robot	0.23	0.400	0.58	3.0	1.9	8.3	0.102	81.0
Villanueva et al. (2011)	Robojelly	0.0542	0.6	0.1	0.5	17.0	314	0.242	1296
Villanueva et al. (2013)	Cyro	0.0847	0.316	0.268	0.12	70.0	826	76.0	11
Wang et al. (2010)	SPC-3 UUV	1.87	1.6	1.2	2.5	194.0	104	n/a	n/a
Wang et al. (2021)	Robotic larval zebrafish	0.133	0.0043	31	83	4.8E-05	3.6E-04	1.535E-06	235
White et al. (2020)	Tunabot Flex	1.17	0.255	4.60	8.0	4.10	3.50	0.190	18.4
White (2022)	Tunabot Prototype	0.64	0.4064	1.6	8.0	10.8	16.9	0.90	18.8
Yu et al. (2016a)	Single-motor-actuated fish	1.14	0.37	3.1	8.0	25.6	22.5	n/a	n/a
Zhong et al. (2017)	Wire-driven robotic fish	0.67	0.31	2.2	3.0	5.6	8.3	0.5	17
Zhu et al. (2019)	Tunabot	1.02	0.255	4.00	14.8	8.67	8.50	0.306	27.8

Why am I here?

Using biology to
inspire new
solution pathways
for fast, efficient
underwater robots



Towards a Mission-
Configurable Stealth
Underwater Batoid

ONR MURI
Program Manager:
Dr R. Brizzolara

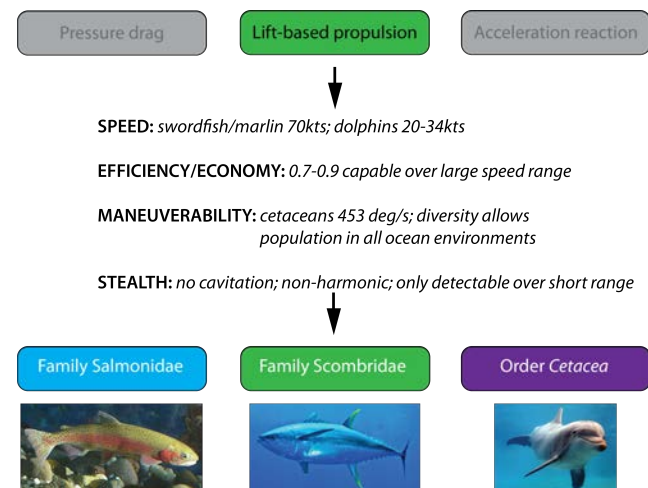
Bio-inspired flexible
propulsors for fast,
efficient swimming:
What physics are we
missing?

ONR MURI
Program Manager:
Dr R. Brizzolara



TUNABOT

High Speed And High Efficiency



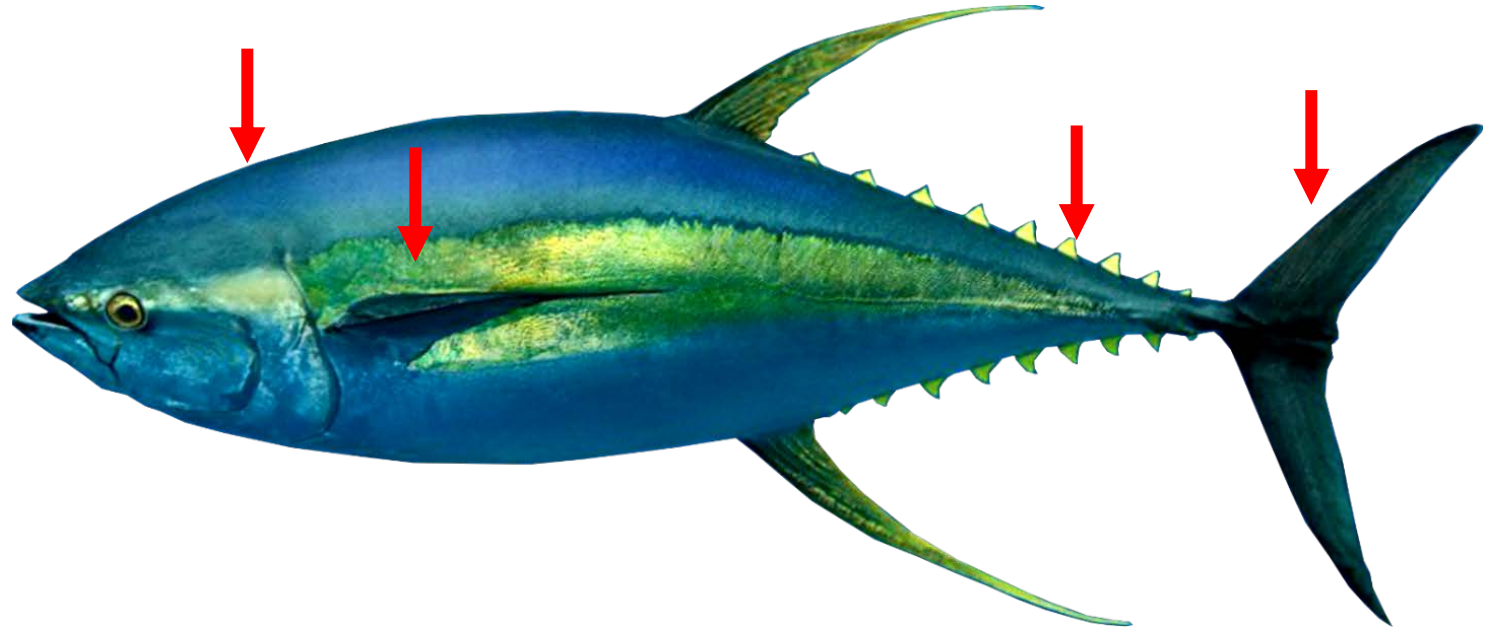
Tuna: biology and anatomy

- Atlantic Bluefin tunas migrate across oceans
- Eastern and western populations mix for feeding but not breeding
- Spawn in Mediterranean and gulf of mexico
- Bluefin grow large (over 1000 lbs) and can sometimes sell for over \$1.5 milion per fish.

Figure 1
At Least 2 Populations of Atlantic Bluefin Tuna: Highly Migratory and Highly Mixed
Western and eastern bluefin mix to feed but separate to breed



Tuna: biology and anatomy



- Key features of tuna:
 - streamlined shape
 - lunate high-aspect ratio tail
 - wing-like pectoral fins
 - finlets
 - caudal peduncle and keel

Research Objectives

1. Study high-performance fish swimming using bio-inspired research platforms
 - Yellowfin tuna (*Thunnus albacares*)

High speed:

Body Lengths per second (BL/s)

High efficiency:

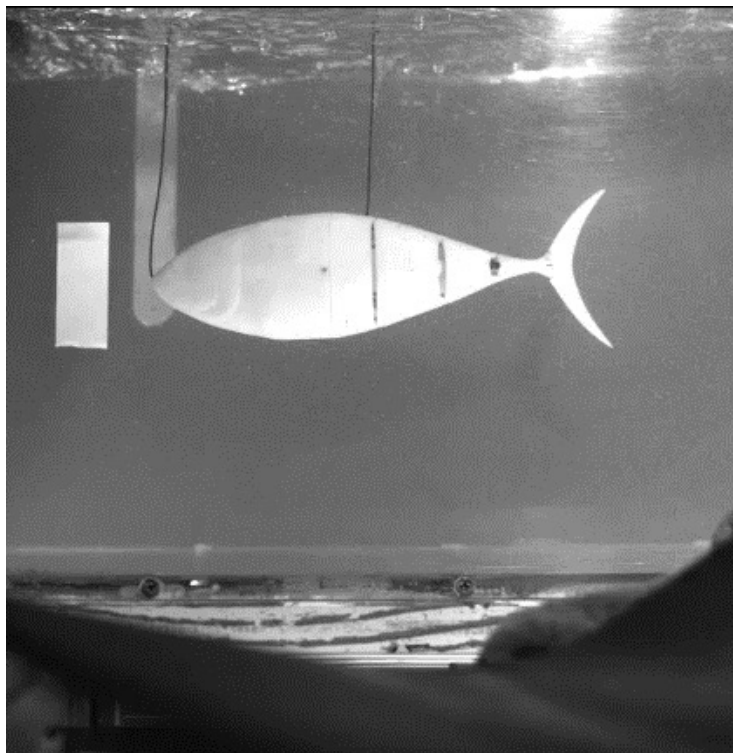
Cost of Transport (J/kg/m)^{COT} = $\frac{\text{Power}}{\text{Mass} \cdot \text{Speed}}$

2. Close the performance gap between biology and robotic systems

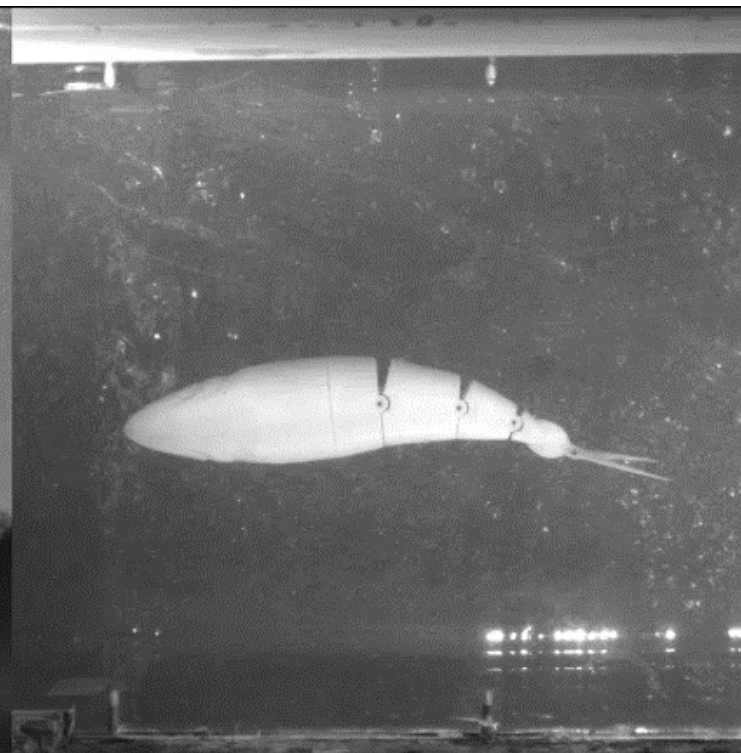


Tunabot Flex Swimming Performance

Lateral View

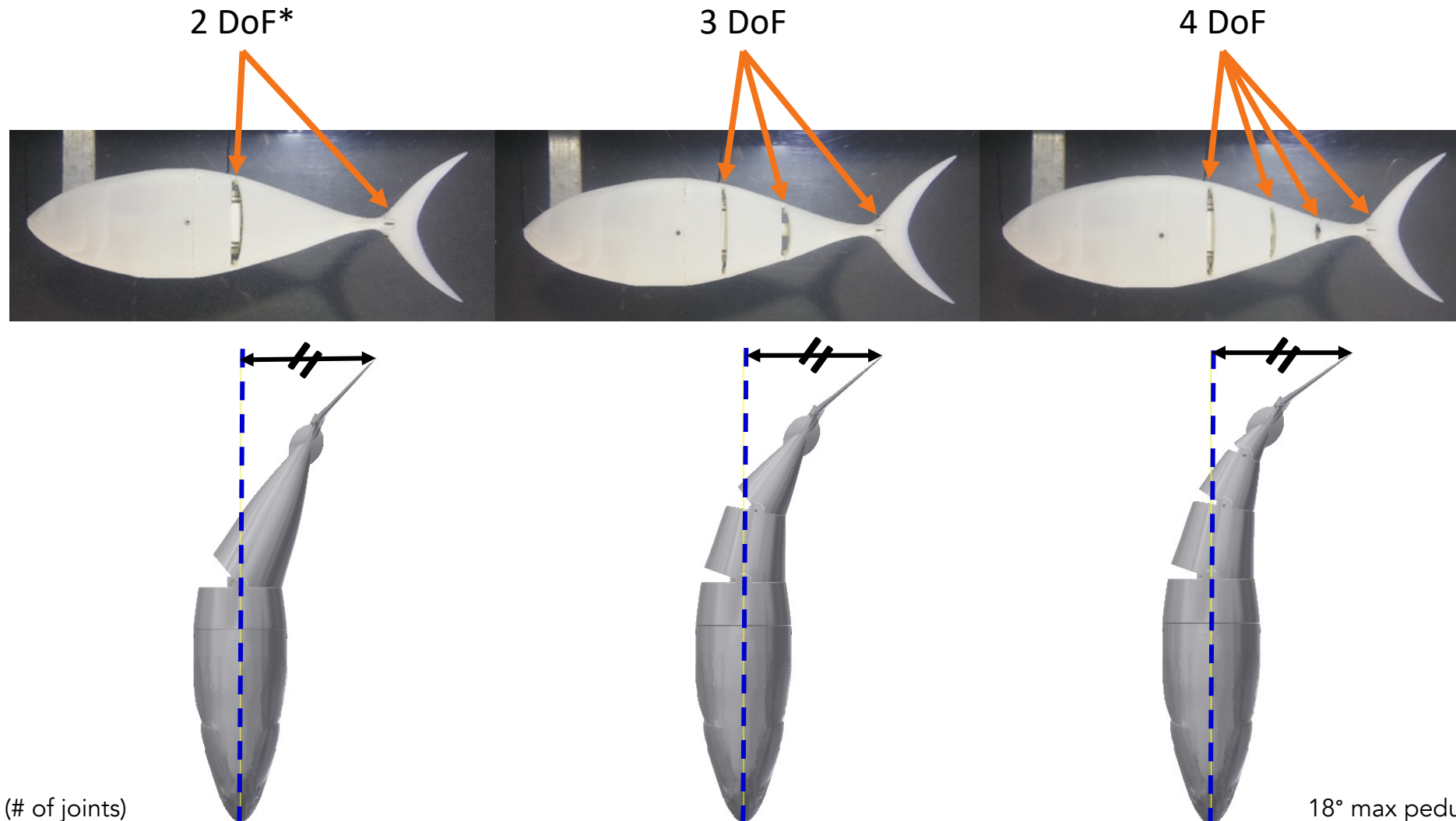


Ventral View



Lauder Lab flow tank, 4.6 BL/s, 8.0 Hz, 0.57x playback speed

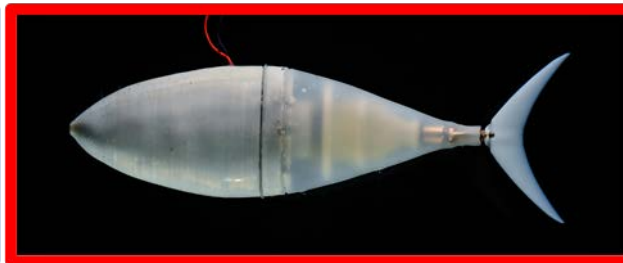
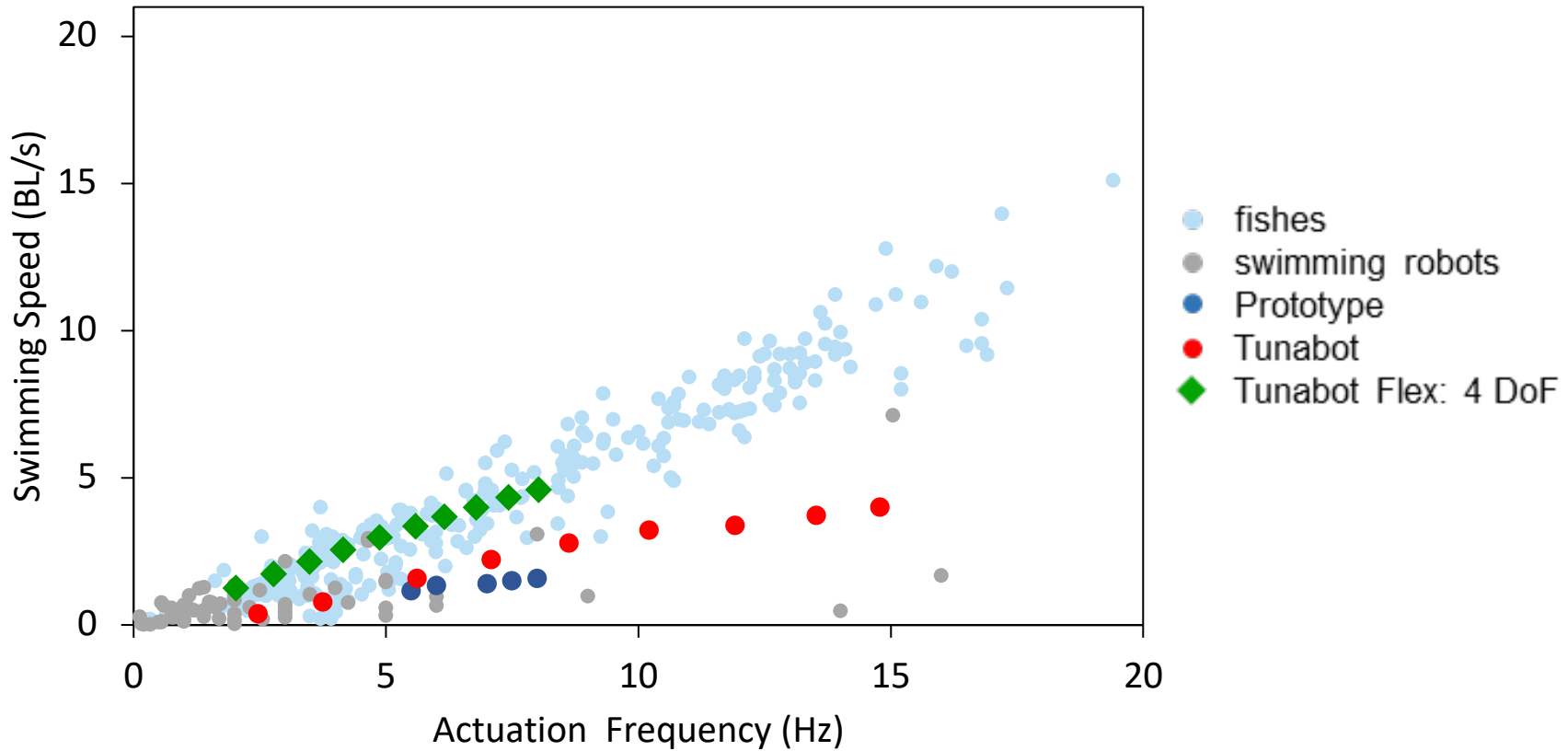
Tunabot Flex Platform Design



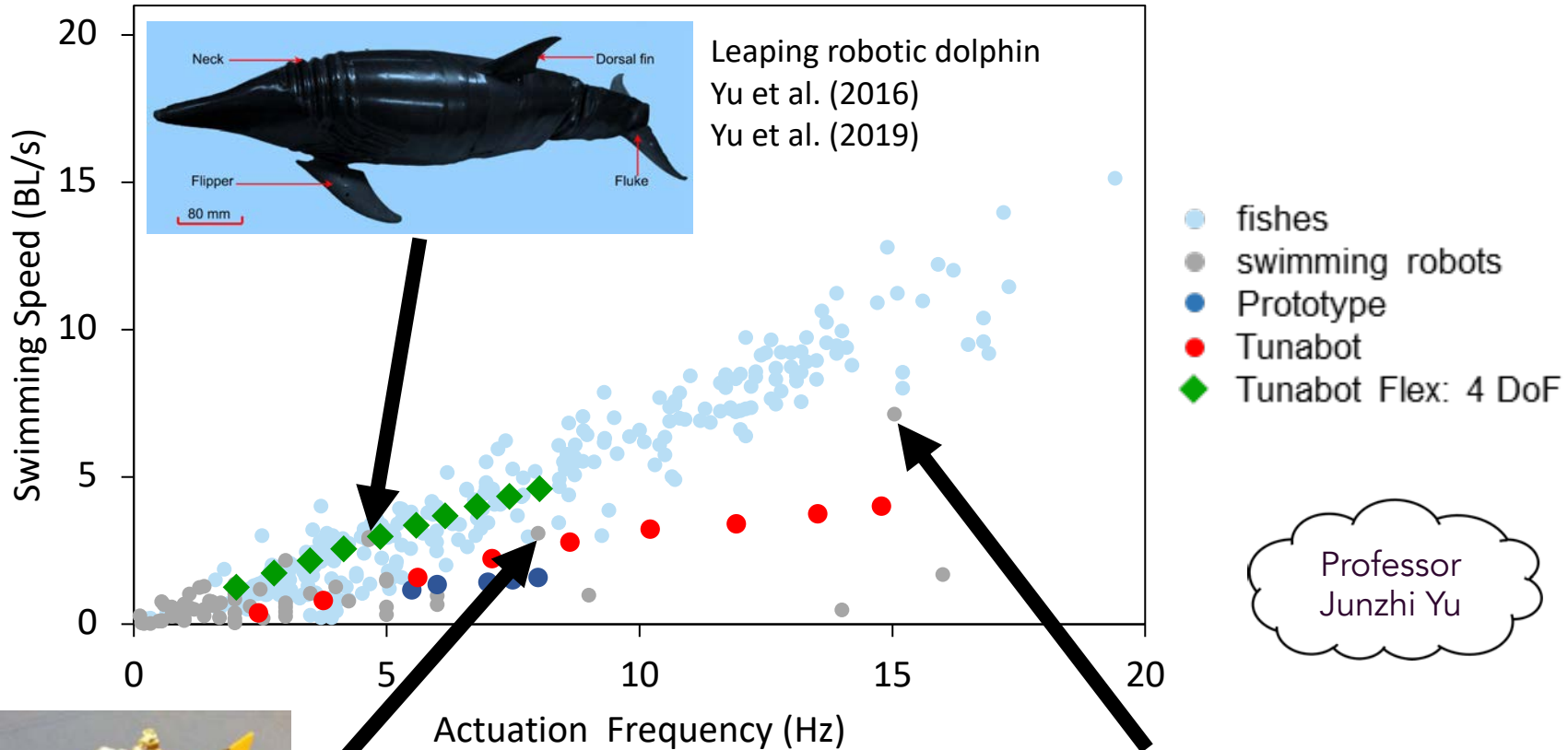
*Degrees of Freedom (# of joints)

18° max peduncle bend angle for all

Performance Space Speed & Frequency



Performance Space Speed & Frequency



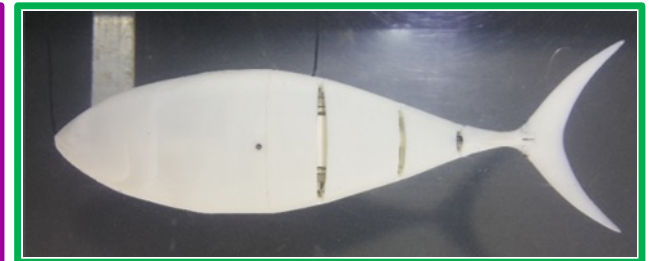
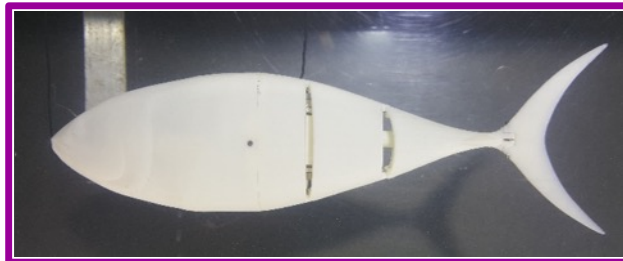
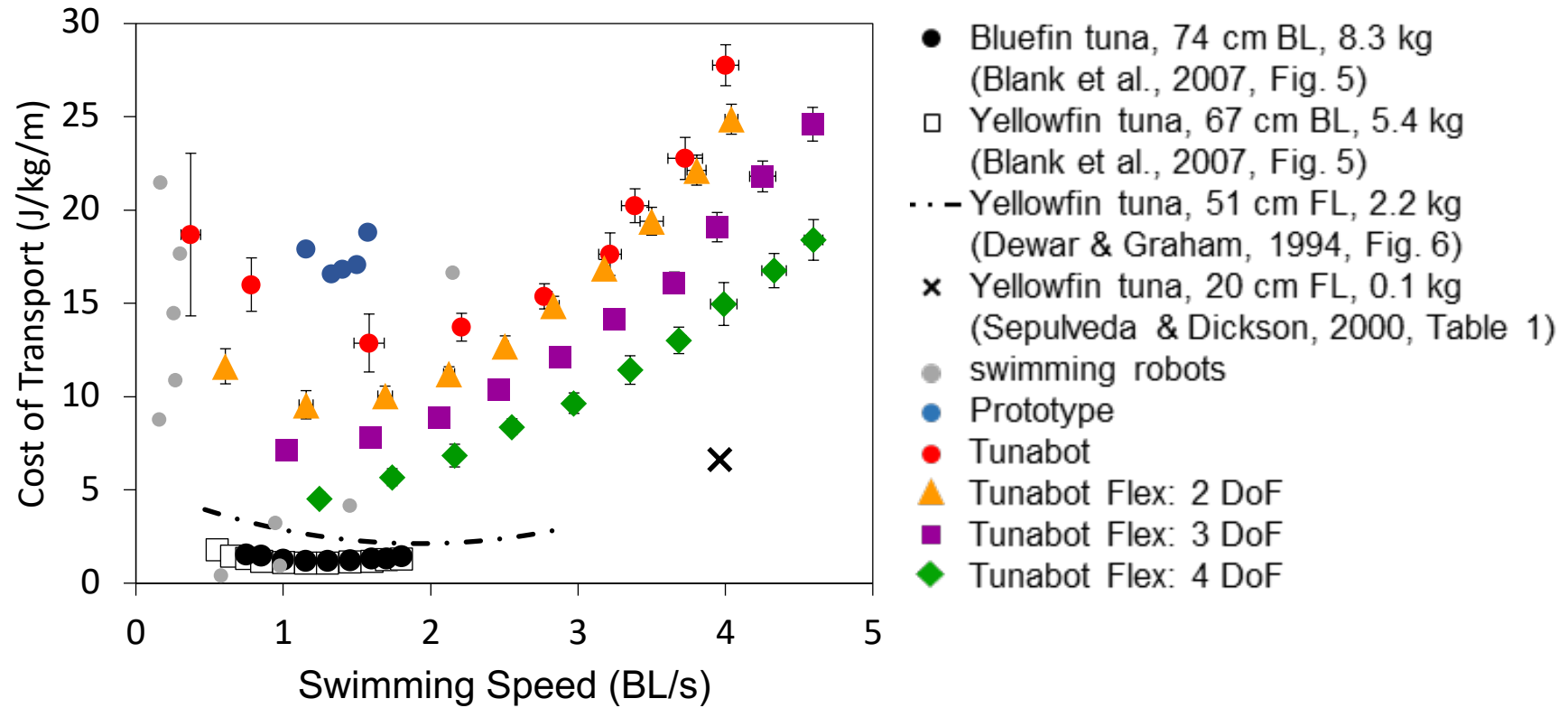
Single-motor-actuated fish
Yu et al. (2016)

Leaping fish bot
Chen et al. (2021)

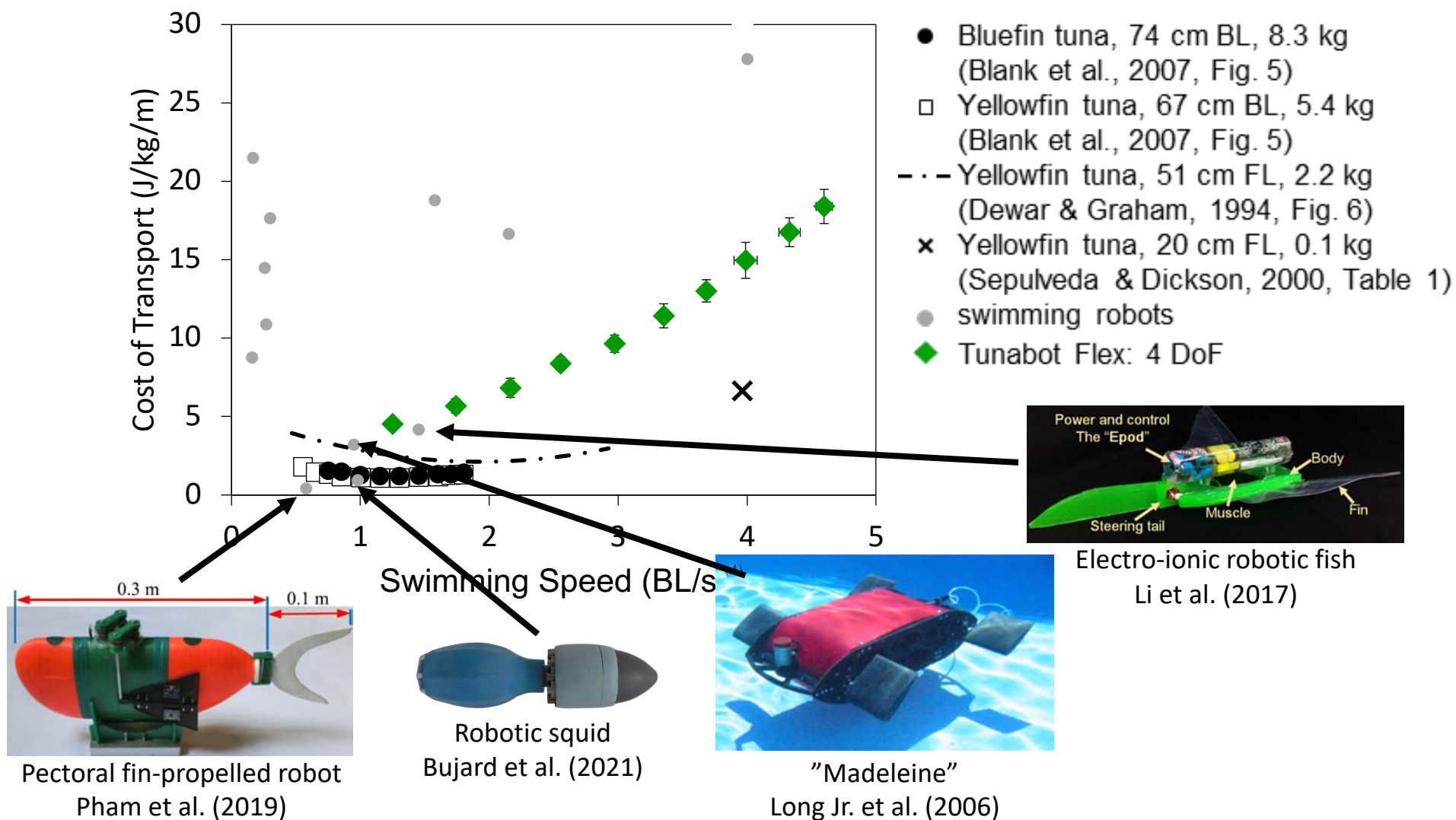


Performance Space

Cost of Transport



Performance Space Cost of Transport



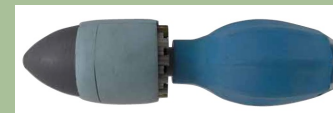
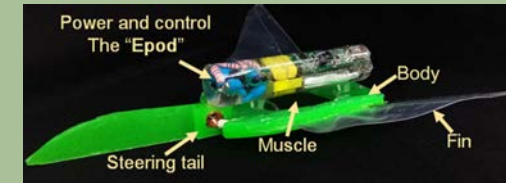
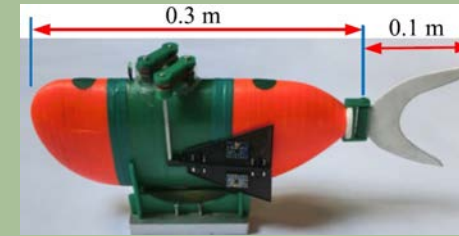
State of the Field

High Speed AND High Efficiency

High Speed

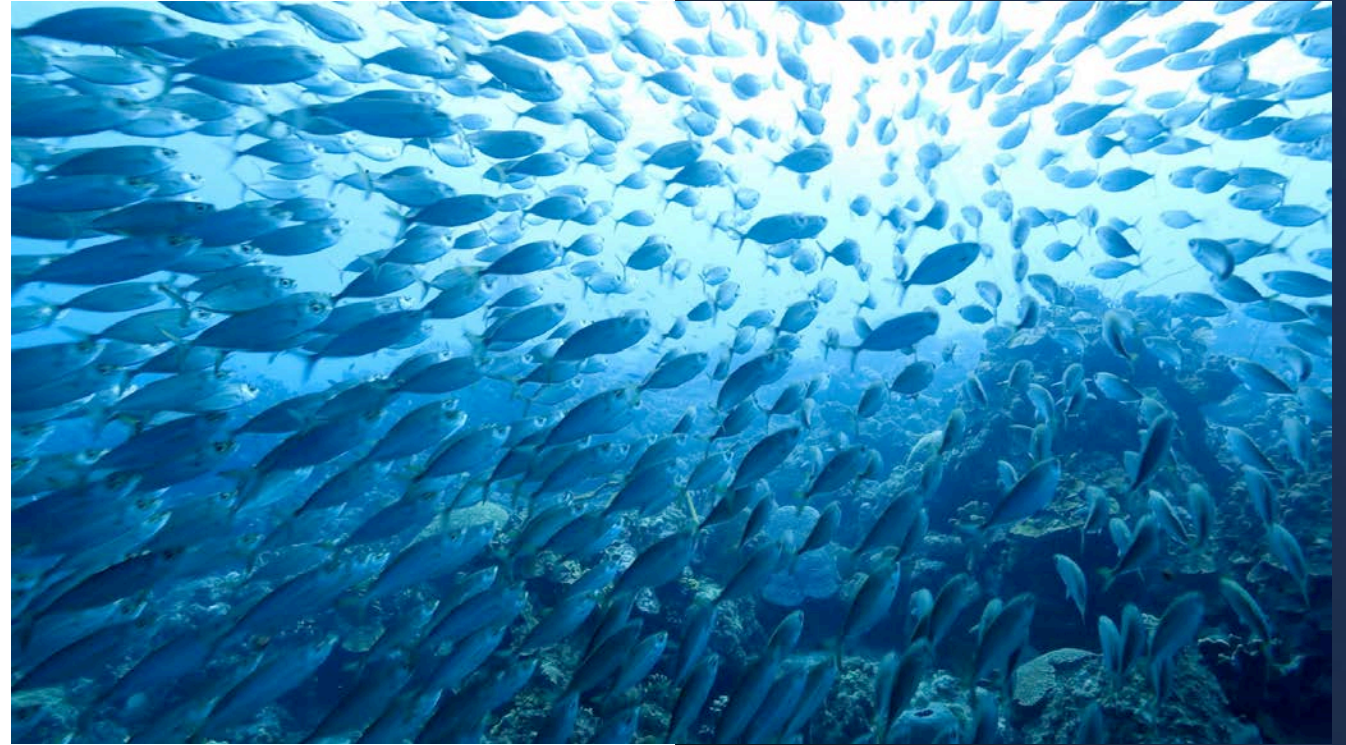


Low Cost of Transport



Fish Schooling

- Simultaneously surveying more area, more intelligently



New MURI: Tentatively starting July 2022

Revealing the Hydrodynamic Principles of Three-Dimensional Fish Schools: From Biology to Schooling Robotics

Principal Investigator

Keith Moored, Lehigh University, kmoored@lehigh.edu



Co-Principal Investigators

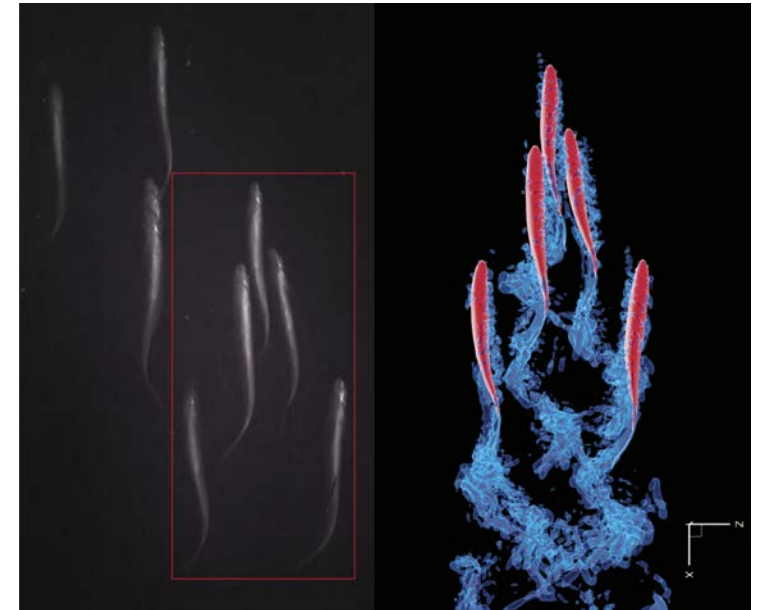
George Lauder, Harvard University

Radhika Nagpal, Princeton University

Hilary Bart-Smith, University of Virginia

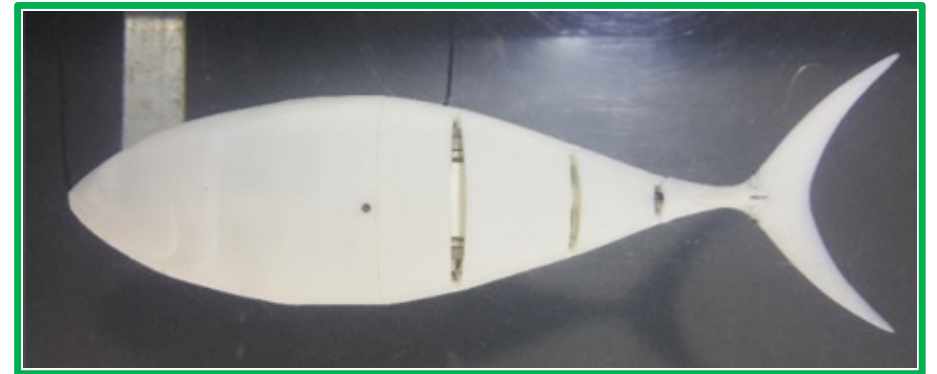
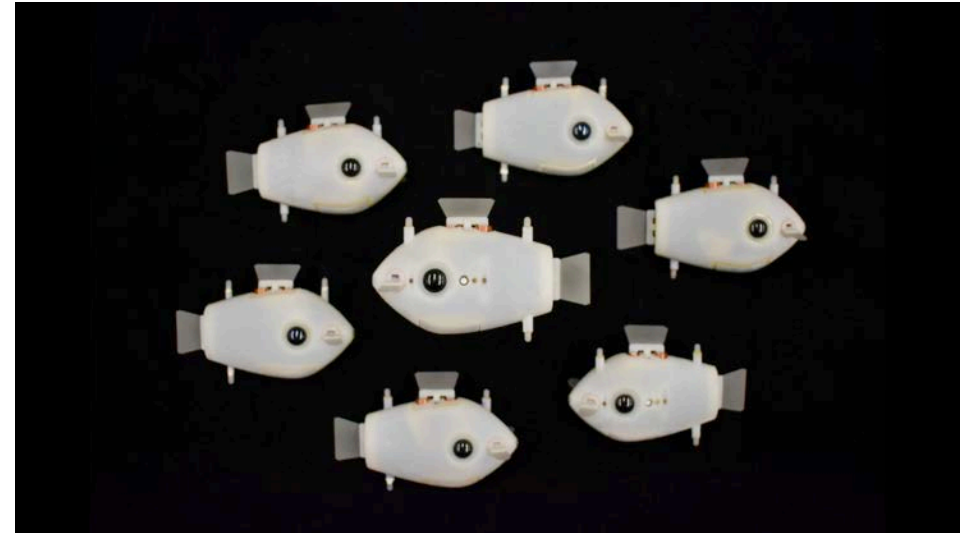
Daniel Quinn, University of Virginia

Haibo Dong, University of Virginia



BlueSwarm: bio-robots with decentralized schooling control

- Fish-inspired robot swarm
- Variable collective behaviors depending on mission requirements
- Berlinger et al. (2021) doi:10.1126/scirobotics.abd866



High-performance + schooling control

FINAL THOUGHTS

- **This is a challenging problem and exciting: lots of questions**
 - Can current technologies be adapted and expanded to explore all oceanic zones?
 - Will we require new technologies and approaches?
 - Is bio-inspired a potential solution path? Soft-systems?
 - Synergies between sensor development and platform and platform control development (control co-design)?
 - Energetics costs?
 - Platform range
 - Recharging implications (Charging stations? On-board power generation?)
 - Economics?